2-Fluoroalkyl A-Ring Analogs of 1,25-Dihydroxyvitamin D₃. Stereocontrolled Total Synthesis via Intramolecular and Intermolecular Diels-Alder Cycloadditions. Preliminary **Biological Testing**

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Intramolecular Diels-Alder (IMDA) cycloadditions of electron-rich trans-vinylic silaketal groups tethered via a chiral, nonracemic 1,3-butanediol auxiliary to electron-poor 2-pyrone-3-carboxylates (e.g. (R)-9, (S)-14) were promoted by zinc dibromide and proceeded unexpectedly in a stepwise, ionic fashion to form exclusively cis-4,5-disubstituted bicyclic lactones 10 and 15 with nearly complete asymmetric induction. Confirmation of the stereochemical outcome of these nonconcerted IMDA cycloadditions was achieved by ¹H NMR spectroscopy and by X-ray crystallography. Fluorinated bicycloadduct (-)-15a was converted smoothly in 13 steps into the lipophilic calcitriol analog 2β -(3'-fluoropropyl)- 1β , 25-dihydroxyvitamin D_3 ((-)-5). Intermolecular, concerted, 11 kbar, inverse-electron-demand 4 + 2-cycloaddition of bis-silylated Z-enol ether 22c, carrying two different silyl groups, with commercial methyl 2-pyrone-3-carboxylate gave in gram amounts only vicinally cis-disubstituted bicycloadduct (\pm)-23c. Chemospecific monodesilylation using sodium azide and then fluorination using Et_2NSF_3 (DAST) gave fluoroalkyl bicyclic lactone (±)-25. This bicyclic lactone (\pm) -25 was transformed into fluorinated, racemic, A-ring phosphine oxide (\pm) -30 that was coupled with enantiomerically pure C,D-ring ketone (+)-21 to form enantiomerically pure diastereomers (-)-4 and (+)-4' as fluorinated, lipophilic, A-ring analogs of 1,25-dihydroxyvitamin D_3 (calcitriol). Preliminary biological testing (Table I) showed that only those diastereomers having the unnatural 1β -hydroxyl group stereochemistry (*i.e.* (+)-3', (+)-4', and (-)-5) had relatively high affinities for the calf thymus vitamin D receptor and significant antiproliferative and differentiationinducing potencies in HL-60 cells.

Searching for new calcitriol analogs like ED-71 (1) and ED-120 (2) for chemotherapy of osteoporosis,¹ we have prepared 2-substituted analog primary alcohol (-)-3 as well as its A-ring stereochemically inverted diastereomer (+)-3', both having significant affinity for the vitamin D binding protein.² As part of our ongoing research program using electronically matched 2-pyrones and various dienophiles for stereocontrolled construction of isolable and versatile bicyclic lactone adducts,³ we report here on the preparation of fluoro analog (-)-4, a fluorinated

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version of (-)-3, as well as on the results of Lewis acidpromoted, intramolecular Diels-Alder (IMDA) cycloadditions of 3-substituted 2-pyrones leading to an asymmetric total synthesis of 2-fluoroalkyl calcitriol analog (-)-5. Preliminary biological evaluations of these seco-



steroids are also presented. During the course of this study, a delicate balance was revealed between concerted and stepwise Diels-Alder cycloadditions.

Results and Discussions

A series of 2-pyrone-3-carboxylate esters with diverse olefinic ester groups was prepared for IMDA cycloadditions.⁴ Allylic and vinylic silanes as well as a vinyl

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4,5-cis-endo-(-)-10b, 15%

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carbonate tethered⁵ to the carboxylate group of 2-pyrone-3-carboxylate all failed to undergo thermal or 11 kbar intramolecular 4 + 2-cycloaddition. In contrast, unsymmetrical enolic silaketals tethered to the pyrone carboxylate group, as in structure (R)-9 (Scheme 1), were successful;⁶ enol silaketals like (R)-9, derived from commercial and enantiomerically pure (R)-1,3-butanediol, were found to undergo IMDA cycloaddition with higher chemical yields and higher stereocontrol than the corresponding enol silaketals derived from enantiomerically pure vicinal diols 1.2-propanediol or 2.3-butanediol. Also, two isopropyl groups attached to the silicon atom (as in silaketal (R)-9) were found to give better results than two methyl groups. Preparation exclusively of the E-geometric isomer enol silaketal 8 from (E)-2-hexenal followed literature precedent (Scheme 1);⁷ ¹H NMR spectroscopy showed a coupling constant of 12 Hz for the two adjacent vinylic hydrogen atoms characteristic of E-double bond stereochemistry.⁷ Lewis acid-promoted⁸ IMDA cycloaddition of pyrone enol silaketal (R)-9 proceeded as shown in Scheme 1 to produce chromatographically separable diastereomers (+)-10a and (-)-10b, isolated in pure form in 74 and 15% yields, respectively. This stereochemical result represents outstanding asymmetric induction by the lone stereogenic carbon center in the (R)-1,3-butanediol chiral auxiliary.

Unambiguously establishing the relative stereochemistry of the bicyclic lactone cycloadducts **10** was essential to understanding the course of this IMDA reaction. Assignment of the 4,5-regiochemistry and of the *cis*-4,5 relative stereochemistry to cycloadducts (+)-**10a** and (-)-**10b** was based on matching the 400 MHz ¹H NMR patterns of chemical shifts and the J_{4,5} coupling constants with those of very close structural analogs, in contrast to the *trans*-4,5-disubstituted isomers, as shown in the accompanying structures.^{2,3} ¹H NMR decoupling experi-



ments were especially useful for these assignments. Within each pair of cis-4,5-disubstituted isomers, the H₄ chemical shifts were very different, but the H₆ chemical shifts were similar; in contrast, the *trans*-4,5-disubstituted isomers had similar H₄ chemical shifts but very different H₆ chemical shifts.

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Scheme 2



Thus, in sharp contrast to concerted intermolecular 4 + 2-cycloadditions of electron-rich enol silyl ethers with electron-poor methyl 2-pyrone-3-carboxylate,^{3,4} this inverse-electron-demand, Lewis acid-promoted, intramolecular 4 + 2-cycloaddition must occur stepwise^{9,10} because the exclusive E-geometry of the dienophile in (R)-9 is lost during formation of only the cis-4,5-disubstituted cycloadducts 10.11 In the absence of zinc dibromide but under 11 kbar pressure, concerted cycloaddition occurred (albeit in low yield) with preservation of dienophile geometry to give mainly the expected trans-4,5-disubstituted cycloadducts. A control experiment showed that these kinetic trans-4,5-disubstituted cycloadducts are stable to the Lewis acid reaction conditions. Another control experiment showed by analytical TLC that an E-configured enol silaketal like 8 (formed via hydrosilvlation of (E)-2-hexenal with *tert*-butyldimethylsilane and chromatographically separated from the corresponding Z-configured enol silaketal) is stable to zinc dibromide in 2/1 toluene/ether even at 0 °C, thereby ruling out the possibility of Lewis acid-promoted trans $\rightarrow cis$ isomerization of enol silaketal (R)-9 before cycloaddition. Apparently therefore in Scheme 1, the Lewis acid polarizes the diene sufficiently to initiate nucleophilic attack by the pendant vinyl silaketal, forming a zwitterionic intermediate containing a siloxy-stabilized carbocation in which the original double bond geometry can be lost.¹⁰ At this time, we do not have a convincing explanation to account for the exclusive formation of only cis-4,5disubstituted cycloadducts 10 rather than a mixture of cis- and trans-4,5-disubstituted cycloadducts via the zwitterionic intermediate. It is noteworthy that the stepwise, ionic nature of this Lewis acid-promoted intramolecular cycloaddition would not have been noticed if only *trans*-4,5-disubstituted cycloadducts had been formed, as originally expected for a concerted cycloaddition starting with the *E*-vinylic silaketal precursor (R)-9.

Establishing the absolute stereochemistry of cycloadducts 10 was more difficult. The four possible *cis*-4,5disubstituted cycloadducts are shown in the accompanying structures. Luckily, the minor cycloadduct 10b was a crystalline solid; X-ray analysis established its absolute stereochemistry to be as shown for 4,5-*cis*-endo-10b. The major cycloadduct could not have 4,5-*cis*-endo stereochemistry because of its nearly zero ¹H NMR J_{4,5} coupling constant; therefore, the major cycloadduct cannot be 4,5*cis*-endo-10c. Thus, the major cycloadduct must be either 4,5-*cis*-exo-10a or 4,5-*cis*-exo-10d. Distinguishing be-



tween diastereomers 10a and 10d is difficult, and the following argument is offered only as a tentative working hypothesis. Molecular models suggest that diastereomers 10a and 10d might be distinguishable from each other by ¹H NMR spectroscopy. Specifically, it appears that the chemical environment of the original stereogenic carbon atom of (R)-1,3-propanediol should be similar for the two R, R, R-diastereomers 10b and 10d. In contrast, (R, R, R)-10b and (R, S, S)-10a should have different ¹H NMR chemical shifts for the methyl doublet and the methine multiplet characteristic of the stereogenic center.

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Indeed, the methyl doublet of (R,S,S)-10a is at 1.40 δ whereas that of (R,R,R)-10b is at 1.34 δ ; likewise, the carbinol hydrogen atom at the stereogenic center in 10a differs in chemical shift from that in **10b** by 0.5 ppm. Therefore, the major cycloadduct was tentatively assigned to have the absolute stereochemistry represented by structure 10a.

To provide a second example of this type of stepwise, Lewis acid-promoted, IMDA cycloaddition and to prepare a 2-fluoroalkyl analog of vitamin D_3 , we selected the enantiomeric chiral auxiliary (S)-1,3-butanediol to form pyrone fluorovinyl silaketal (S)-14 (Scheme 2). The requisite fluorovinyloxysilyl chloride 13 was prepared by hydrosilylation⁷ of 5-fluoro-(E)-2-pentenal, itself formed in several steps according to literature precedent from epichlorohydrin (Scheme 2).¹² Although fluorovinyl silaketal (S)-14 was considerably less stable than its nonfluorinated analog (R)-9, when used immediately after its preparation, it underwent a similar zinc dibromide-promoted IMDA cycloaddition to give easily separable cis-4,5-disubstituted cycloadducts (-)-15a and (-)-15b in 46 and 12% yields, respectively. The 400 MHz ¹H NMR spectra of these cycloadducts very closely matched those of the cis-4,5-disubstituted cycloadducts (+)-10a and (-)-10b (Scheme 1). Using the (S)-1,3butanediol chiral auxiliary to prepare pyrone vinyl silaketal (S)-14 gave the cycloadducts 15 that are enantiomeric to cycloadducts 10.

Using other Lewis acids⁸ (e.g. MgBr₂, Et₂AlCl, TiCl₄, LiClO₄) to promote the IMDA cycloaddition of fluorovinyl silaketal (S)-14 was successful but offered no advantage over zinc dibromide in the yield of cis-4,5-disubstituted cycloadducts (-)-15a and (-)-15b or in the ratio of these cycloadducts.

Diastereomerically pure fluorinated cycloadduct (-)-15a was carried on toward a 2-fluoroalkyl-1,25-dihydroxyvitamin D_3 analog. Opening of the silaketal ring using HF followed immediately by silvlation gave O-protected bicyclic lactone 16 (Scheme 3). Opening of the lactone ring chemospecifically with a large excess of lithium allyl oxide gave an intermediate mixed allyl methyl malonate (isolated when only 1 equiv of allyl oxide was used) that, unexpectedly, underwent spontaneous decarboxylation¹³ with concomitant conjugation of the olefinic double bond to the ester group to form stereospecifically tetrasubstituted cyclohexene (-)-17. After O-silylation, reduction of the conjugated enoate ester led to allylic alcohol (-)-18 that underwent smooth, one-flask two-carbon homologation via a Claisen rearrangement followed by a spontaneous thermal sulfoxide elimination using our sulfinylated orthoester protocol;¹⁴ photochemical isomerization¹⁵ of $E \rightarrow Z$ -dienoate gave A-ring chiron (-)-19. Established reactions as shown in Scheme 3 provided the crucial, fully protected, enantiomerically pure, A-ring phosphine oxide (+)-20.² Lythgoe-type coupling¹⁶ of the conjugate base of this phosphine oxide, generated using



phenyllithium,¹⁷ with C,D-ring ketone (+)-21 of natural absolute configuration¹⁵ produced an O-silylated product that underwent fluoride-promoted desilylation to form a fluorinated, lipophilic vitamin D analog with tentative stereochemical assignment as 2β -(3'-fluoropropyl)-1 β ,25dihydroxyvitamin D_3 analog (-)-5 having the unnatural 1β -hydroxyl group orientation.^{18,19}

Having in hand the various synthetic intermediates leading up to 2-hydroxybutyl analog (-)-3,² we explored when it would be easiest to replace the primary alcohol

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group by a fluorine atom,²⁰ leading to preparation of fluorinated vitamin D analog (-)-4. Toward this end, failed attempts included treating primary-secondary bissilyl ether cycloadduct 23a with tetra-n-butylammonium fluoride,²¹ aiming for chemoselective deprotection of the sterically more accessible primary alcohol, and treating differently protected bis-silvl ether cycloadduct 23b with sodium hydride/HMPA, a procedure reported to selectively desilylate tert-butyldiphenylsilyl ethers in the presence of tert-butyldimethylsilyl ethers.²² Changing the primary alcohol protecting group to a diphenylmethylsilyl ether in the reactant enol silyl ether 22 produced cycloadduct (\pm) -23c in which somewhat capricious hydrolytic deprotection on silica gel formed free primary alcohol (\pm) -24. More reliable chemospecific deprotection of this primary silyl ether group in bis-silyl ether cycloadduct (\pm) -23c was finally achieved using sodium azide (Scheme 4).23 Although the primary triflate corresponding to alcohol (\pm) -24 failed to undergo nucleophilic displacement with fluoride ion, direct fluorination of primary alcohol (\pm) -24 with Et₂NSF₃ (DAST) was successful (Scheme 4).²⁴ It should be noted that several attempts for converting 6-fluorohexanal into the corresponding enol silyl ether²⁵ for cycloaddition with methyl 2-pyrone-3-carboxylate as a direct approach to fluorinated cycloadduct (\pm) -25 failed. Finally, it is worth emphasizing that the gram-scale, concerted cycloaddition of bissilyl Z-enol ether 22c led, as expected on the basis of a recent close analogy,3 exclusively to vicinally cis-disubstituted bicyclic lactone (\pm) -23c in 62% yield, with starting commercial methyl 2-pyrone-3-carboxylate recovered in 15-20% yield.

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Also in accord with recent precedent, fluorinated bicyclic lactone (\pm) -25 was converted into racemic, fluorinated, A-ring phosphine oxide (\pm) -30 (Scheme 5).² Noteworthy features of the transformations in Scheme 5 are as follows: (1) chemospecific lactone opening by lithium allyl oxide² to form cyclohexene (\pm) -26 even in the presence of other electrophilic centers such as the methyl ester, the silyl ether, and the primary fluoralkyl group; (2) mild and non-high pressure deallyloxycarboxylation²⁶ with concomitant conjugation of the enoate double bond to form cyclohexene (\pm) -27; (3) survival of the primary fluoroalkyl group to i-Bu₂AlH reductions in preparation of alcohol (\pm) -28 and of phosphine oxide (\pm) -30; and (4) stereoselective generation of the desired Z-dienoate (\pm) -30, easily separated chromatographically from its *E*-isomer, thereby avoiding the need for photochemical isomerization,¹⁵ by one-pot two-carbon homologation using a phenylsulfinyl orthoester.¹⁴ Coupling² of the conjugate base of racemic allylic phosphine oxide (\pm) -**30** with the enantiomerically pure C,D-ring ketone (+)-21 gave HPLC-separable, enantiomerically pure fluoroalkyl seco-steroids (-)-4 and (+)-4'.

Tentative assignment of A-ring stereochemistry to these fluoroalkyl A-ring analogs (-)-4 and (+)-4' was made by comparing the characteristic portions of their ¹H NMR spectra with those of hydroxyalkyl A-ring analog (-)-3 and its diastereomer (+)-3' (corresponding to (+)-4') as well as their characteristic signs of optical rotation.^{2,27} Specifically, the chemical shifts of the C₁₈-methyl group (δ 0.55 vs 0.53) and the C₁₉-methylene group (δ 5.01 vs 4.99) were diagnostic. The ¹⁹F NMR spectra of diastereomers (-)-4 and (+)-4' confirmed the presence of a primary alkyl fluoride group.²⁸ These steroidal alkyl fluorides were much more soluble in organic solvents (*i.e.* more lipophilic) and much less polar on TLC than were the corresponding alcohols (-)-3 and (+)-3'.

Replacing the primary hydroxyl group in analog (-)-3 with a fluorine atom and inverting the stereochemistry at positions 1 and 2 (*i.e.* analog (-)-5) caused an attenuation in the relative binding affinity to the vitamin D binding protein (DBP) but an increase in the relative binding affinity to the calf thymus vitamin D receptor (VDR), as specified in Table 1, in which the data were generated in vitro according to the previously described protocol.^{1,29} The relative in vitro potencies for inhibition of human leukemic HL-60 cell proliferation and induction of HL-60 cell differentiation as well as the relative VDR binding affinities were higher for the unnatural 1β hydroxyl diastereomers (*i.e.* (+)-3' vs (-)-3 and (+)-4' vs (-)-4). The strong similarity in the relative binding affinities of analogs (+)-3', (+)-4', and (-)-5 as well as their similar activities in HL-60 cells support the absolute stereochemistry of (-)-5 being the same as that in analogs (+)-3' and (+)-4' at the crucial 1- and 2-positions. The lipophilic fluorinated analogs (+)-4' and (-)-5 are expected to have in vivo pharmacodynamics and transport characteristics different than those of osteoporosis

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 Table 1. Chemical Structure-Biological Activity

 Relationships

	relative binding affinities		inhibition of cell	induction of cell
	DBP	VDR	proliferation	differentiation
1,25-(OH) ₂ D ₃	1	1	1	1
ED-71	3.7	0.21		
ED-120	1.8	0.01		
(-) -3	5.0	0.01	< 0.1	< 0.1
(+) -3 ′	1.9	0.28	0.58	0.84
(-)-4	1.2	0.001	< 0.1	< 0.1
(+) -4 ′	0.40	0.16	0.50	0.23
(-)-5	0.83	0.10	0.20	0.41

drug candidate ED-71 (1), and, therefore, they deserve further *in vivo* biological evaluation to determine whether they have any practical value in preventing or curing osteoporosis.

The structure-activity relationships described here, along with those reported recently,^{1-3,19} emphasize that considerable structural and stereochemical changes at the 1-3-positions of calcitriol can be made while still maintaining significant regulation of calcium metabolism and cell differentiation. These combined results, therefore, encourage design, synthesis, and biological evaluation of additional A-ring analogs of calcitriol that may have practical value in preventing or diminishing the bone mineral loss characteristic of osteoporosis and in restoring desirable differentiating ability to nondifferentiating cells.

Experimental Section¹⁴

The purity of most products was judged to be >95% on the basis of their chromatographic homogeneity. The only exceptions were the final coupling products in which the purity was somewhat lower due to the small amounts of seco-steroids produced, making their purification and spectroscopic characterization difficult.

Pyrone Ester (**R**)-7. A mixture of hexamethyldisilane (4.75 g, 32.4 mmol) and iodine (8.23 g, 32.4 mmol) was heated at 65 °C in a dry 500 mL round-bottomed flask equipped with a reservoir and a long reflux condenser. A violent exothermic reaction occurred, and a homogeneous reddish brown solution resulted, which was heated under reflux for 1.5 h to form a colorless liquid (hexamethyldisilane was quantitatively converted to iodotrimethylsilane). Methyl 2-oxo-2H-pyran-3-carboxylate (5.0 g, 32.4 mmol) in 50 mL of CHCl₃ was added, and the mixture heated at reflux until the reaction was complete by TLC analysis (*ca*. 25 h). The reaction mixture was cooled to 25 °C, and 2 mL of water was added. The

mixture was strirred for 10 min and then dissolved with CH₂-Cl₂ (100 mL). Saturated aqueous sodium thiosulfate (5 mL) was added, and the mixture was stirred until complete decoloration was evident. The organic layer was separated, and the aqueous layer was extracted with CH₂Cl₂ (3 × 100 mL). The combined organic solution was then washed with saturated sodium thiosulfate solution (1 × 20 mL), dried (MgSO₄), filtered, and evaporated under reduced pressure, and the residue was dried using a vacuum pump. The 2-pyrone-3-carboxylic acid was recrystallized from toluene to afford 3.53 g (78% yield) as yellow crystals: mp 121–123 °C; ¹H NMR (CDCl₃) δ 8.56 (dd, J = 2.2, 6.5 Hz, 1H), 7.83 (dd, J = 2.2, 4.9 Hz, 1H), 6.68 (dd, J = 5.1, 6.9 Hz, 1H); IR (CHCl₃) 1749, 1690, 1555, 1414, 1390 cm⁻¹. Anal. Calcd for C₆H₄O₄: C, 51.44; H, 2.91. Found: C, 51.47; H, 2.89.

To 2.0 g (14.3 mmol) of this carboxylic acid in benzene (50 mL) was added oxalyl chloride (8 mL, 91.4 mmol). A few drops of DMF were added which initiated a vigorous reaction. The reaction mixture was stirred at rt for 2 h. The solvent was evaporated under reduced pressure. The residue was redissolved in benzene and evaporated again. The residue was dried under a vacuum pump and recrystallized from benzene to afford 2.26 g (100% yield) of the corresponding acid chloride as yellow crystals: mp 98–100 °C; ¹H NMR (CDCl₃) δ 8.51 (dd, J = 2.22, 7.08 Hz, 1H), 7.82 (dd, J = 2.24, 5.06 Hz, 1H), 6.51 (dd, J = 4.98, 7.05 Hz, 1H); IR (CHCl₃) 3013, 1813, 1784, 1743, 1625, 1543 cm⁻¹. Anal. Calcd for C₆H₄ClO₃: C, 45.45; H, 1.91; Cl, 22.39. Found: C, 45.61; H, 1.96; Cl, 22.24.

To 0.919 g (10.2 mmol) of (*R*)-1,3-butanediol, 1.414 mL (19.0 mmol) of triethylamine, 0.257 g (2 mmol) of 4-(dimethylamino)pyridine (DMAP), and 20 mL of CH₂Cl₂ was added 1.543 g (10.2 mmol) of *tert*-butyldimethylsilyl chloride (TBDMSCl) dissolved in 10 mL of CH₂Cl₂ at -78 °C. Upon addition, the reaction mixture was allowed to warm to rt and stirred for 27 h. The reaction mixture was then diluted with 100 mL of ether, washed with H₂O and saturated NH₄Cl, dried over MgSO₄, concentrated *in vacuo*, and purified by column chromatography (silica gel, 80/20 hexane/EtOAc) to afford 1.789 g of the monoprotected diol (+)-**6** in 87% yield: ¹H NMR (CDCl₃) δ 4.02–3.94 (m, 1H), 3.90–3.83 (m, 1H), 3.80–3.74 (m, 1H), 1.69–1.55 (m, 2H), 1.16 (d, J = 6.4 Hz, 3H), 0.87 (s, 9H), 0.04 (bs, 6H); ¹³C NMR (CDCl₃) δ 68.1, 25.8, 23.3, 18.1, -5.57, -5.62; FT-IR (CHCl₃) 3482, 2956, 2859, 1472, 1260 cm⁻¹.

To 1.85 g (9.1 mmol) of the monosilylated diol (R)-6 in 20 mL of anhydrous CH₂Cl₂ were added 1.44 g (9.28 mmol) of 2-pyrone-3-carboxylic acid chloride in 20 mL of CH₂Cl₂ and 1.58 mL (11.3 mmol) of triethylamine at -78 °C. The reaction mixture was then slowly warmed to rt over 2 h, washed with H₂O, dried over MgSO₄, concentrated in vacuo, and purified by column chromatography (80/20 hexane/EtOAc) to give 1.89 g of the O-silylated derivative of pyrone ester (R)-7 as a brown oil in 64% yield: ¹H NMR (CDCl₃) δ 8.10 (dd, J = 6.4, 2.4 Hz, 1H), 7.66 (dd, J = 5.2, 2.4 Hz, 1H), 5.24–5.17 (m, 1H), 3.74– 3.64 (m, 2H), 1.95 - 1.86 (m, 1H), 1.81 - 1.73 (m, 1H), 1.32 (d, J= 6.4, 3.0 Hz, 3H, 0.84 (s, 9H), -0.01 (s, 3H), -0.02 (s, 3H); $^{13}\mathrm{C}$ NMR (CDCl_3) δ 162.5, 157.1, 156.1, 147.8, 118.4, 105.6, 70.2, 59.2, 38.7, 25.8, 20.0, 18.1, -5.5, -5.6; FT-IR (CHCl₃) 2930, 2927, 2857, 1758, 1705, 1552, 1463, 1362, 1292, 1259, 1244 cm⁻¹; HRMS m/e (M⁺ - t-Bu) calcd for C₁₂H₁₇O₅Si 269.0845, found 269.0849.

To 1.89 g (5.8 mmol) of this O-silylated derivative of (R)-7 was added 30 mL of 5% HF in acetronitrile at rt. After 8 min, the reaction mixture was neutralized with aqueous NaHCO₃, extracted with CHCl₃ (2 × 100 mL), dried over MgSO₄, concentrated *in vacuo*, and purified by column chromatography (100% EtOAc) to afford 0.75 g of the alcohol (+)-7 in 61% yield as a yellow oil. The alcohol (R)-7 was used for the next reaction without a complete characterization due to its instability.

Enol Silaketal (R)-9. Into a 20 mL hydrolysis tube were placed 0.726 g (7.4 mmol) of *trans*-2-hexenal, 1.155 g (7.6 mmol) of chlorodiisopropylsilane, and 0.033 g of tris(triphenylphosphine)rhodium chloride. The tube was then sealed, stirred for 10 h at rt, and heated at 90 °C for 14 h. The reaction mixture was decanted and directly purified by a bulb to bulb distillation (60–70 °C at 0.5 mmHg) to afford 1.380 g of the enol silyl chloride **8** as a colorless liquid in 75% yield,

contaminated with a trace amount of impurities: ¹H NMR (400 MHz, CDCl₃) δ 6.30 (dt, J = 12.0, 1.6 Hz, 1H), 5.13 (dt, J = 12.0, 7.6 Hz, 1H), 1.93–1.87 (m, 2H), 1.34–1.28 (m, 4H), 1.14–1.10 (m, 2H), 1.09 (d, J = 4.8 Hz, 6H), 1.08 (d, J = 4.8 Hz, 6H), 0.91–0.86 (m, 3H); ¹³C NMR (400 MHz, CDCl₃) δ 133.4, 113.4, 32.3, 26.8, 22.1, 16.6, 16.5, 16.3, 14.8, 13.9. Because of its lability, this material was carried on immediately.

To 71.3 mg (0.3 mmol) of the alcohol (R)-7 and 10 mL of anhydrous CH₂Cl₂ were added 100.4 mg (0.4 mmol) of silyl chloride 8 in 10 mL of anhydrous CH_2Cl_2 and 0.05 mL (0.3 mmol) of triethylamine at -78 °C. Upon addition, the reaction mixture was allowed to warm to rt and stirred for 27 h. The reaction mixture was slowly warmed to rt over 7 h, diluted with 50 mL of ether, washed with H₂O and brine, dried over MgSO₄, concentrated in vacuo, and purified by column chromatography (20/80 EtOAc/hexane) to give 97.3 mg of the pyrone enol silaketal (R)-9 in 67% yield as a greenish yellow oil (slowly decomposing): ¹H NMR (CDCl₃) δ 8.13 (dd, J = 6.8, 2.4 Hz, 1H), 7.66 (dd, J = 5.1, 2.3 Hz, 1H), 6.33 (dd, J = 6.8, 5.0 Hz, 1H), 6.30 (dt, J = 11.8, 1.4 Hz, 1H), 3.91-3.86 (m, 2H), 2.04–1.85 (m, 1H), 1.88–1.80 (m, 2H), 1.37 (d, J = 6.2Hz, 3 H), 1.31-1.25 (m, 4H), 1.03 (d, J = 3.6 Hz, 12H), 1.03-1.00 (m, 1H, overlapped with doublet at δ 1.03), 0.89–0.85 (m, 3H). Because of its instability, silaketal (R)-9 was carried on immediately.

Cycloadducts (+)-10a and (-)-10b. To a flame-dried 50 mL flask charged with 0.310 g (7.3 mmol) of the pyrone enol ether (R)-9, 3 mL of anhydrous ether, and 6 mL of anhydrous toluene was added 0.160 g (7.3 mmol) of ZnBr_2 . Upon addition, the reaction mixture was cooled to -30 °C and stirred for 4 days. The reaction mixture was then concentrated by rotary evaporator and directly purified by column chromatography (97/3 hexane/EtOAc) to afford 0.230 g of the cycloadduct (+)-10a in 74% yield along with 0.045 g (15% yield) of the crystalline cycloadduct (-)-10b. For cycloadduct (+)-10a: ^{1}H NMR (CDCl₃) δ 6.77 (dt, J = 8.0, 1.2 Hz, 1H), 6.59 (dt, J =8.0, 5.2 Hz, 1H), 5.54–5.46 (m, 1H), 4.98 (d, J = 5.2, 1.6 Hz, 1H), 4.38 (s, 1H), 3.89-3.77 (m, 2H), 1.88-1.28 (m, 8H), 1.324(d, J = 6.4 Hz, 3H), 1.25-1.16 (m, 1H), 1.03 (d, J = 3.2 Hz,3H), 1.01 (d, J = 3.6 Hz, 3H), 0.92–0.87 (m, 9H); ¹³C NMR $(CDCl_3) \delta 168.5, 167.0, 130.8, 129.3, 76.4, 75.4, 70.3, 62.6, 59.8,$ 49.2, 37.7, 30.1, 29.3, 22.4, 20.6, 17.6, 17.4, 17.3, 17.2, 13.8, 12.7, 10.5; FT-IR (CHCl₃) 2945, 2868, 1769, 1724, 1464, 1364, 1291, 1259 cm⁻¹; HRMS m/e (M⁺ – *i*-Pr) calcd for C₁₉H₃₁O₆Si 381.1733, found 381.1740; $[\alpha]^{24}$ 17.5 ($c = 22 \text{ mg/mL}, \text{CH}_2\text{Cl}_2$) For cycloadduct (–)-10b: ¹H NMR (CDCl₃) δ 6.83 (dt, J = 8.0, 1.2 Hz, 1H, 6.51 (dd, J = 8.0, 5.0 Hz, 1H), 5.56-5.60 (m, 1H), $5.06 \text{ (m, 1H)}, 4.94 \text{ (d, } J = 7.6 \text{ Hz}, 1\text{H}), 3.95 \text{ (m, 1H)}, 3.84 \text{$ 1H), 2.41 (m, 2H), 1.85 (m, 2H), 1.5-1.2 (m, 9H), 1.40 (d, J =6.4 Hz, 3H); HRMS m/e (M⁺ - *i*-Pr) calcd for C₁₉H₁₃O₆Si 381.1733, found 381.1739; $[\alpha]^{24}$ _D -1.8 (c = 35 mg/mL, CH₂-Cl₂). Single crystal X-ray analysis showed the absolute stereochemistry of (-)-10b to be as shown in Scheme 1.

1,3-Bis(methylthio)-2-methoxypropane (11): colorless oil; ¹H NMR δ 3.49 (m, 1H), 3.41 (s, 3H), 2.73 (m, 4H), 2.14 (s, 6H); data consistent with literature data.¹²

trans-1,3-Bis(methylthio)-5-fluoro-1-pentene (12). To diisopropylamine (5.11 g, 50.5 mmol) in THF (100 mL) was added n-BuLi (1.5 M in hexanes, 7.1 mL, 50.5 mmol) at -78 °C. After 10 min, 1,3-bis(methylthio)-2-methoxypropane (11, 4.2 g) in THF (20 mL) was added *via* cannula at -78 °C. After addition was completed, the reaction mixture was warmed to 0 °C (the reaction mixture changed from dark brown to a deep purple solution) and stirred for 2.5 h. The resulting purple solution was cooled to -78 °C, and 1-bromo-2-fluoroethane (3.53 g, 27.8 mmol) was added dropwise. The reaction mixture was warmed to rt, after which time reaction was complete by analytical TLC (ca. 4 h). The reaction mixture was diluted with ether and washed successively with aqueous NH₄Cl, water, and brine. The combined aqueous washes were extracted with ether $(2 \times 100 \text{ mL})$. The combined ethereal solution was dried $(MgSO_4)$, filtered, and evaporated under reduced pressure. The residue was subjected to silica gel column chromatography (eluent, 95/5 hexanes/ethyl acetate) to give an inseparable trans/cis mixture of 1,3-bis(methylthio)-5-fluoro-1-pentene (12) (4.67 g, 100%) as a deep orange oil: R_f 0.62 (70/30 hexane/ethyl acetate); ¹H NMR (CDCl₃) δ 6.12 (d, J = 14.8 Hz, 1H), 5.16 (dd, J = 9.6, 14.6 Hz, 1H), 4.63– 4.41 (m, 2H), 3.35 (m, 1H), 2.27 (s, 3H), 2.01 (s, 3H); ¹³C NMR (CDCl₃) δ 126.8, 124.9, 81.1 (d, J = 658.4 Hz), 45.7 (d, J = 18Hz), 35.2 (d, J = 78.8 Hz), 14.7 (d, J = 13.8 Hz); IR (CHCl₃) 2978, 2906, 1596, 1467, 1425, 1390, 1220 cm⁻¹; HRMS m/e(M⁺) calcd for C₇H₁₃FS₂ 180.0443, found 180.0444.

Enol Silyl Chloride 13. To a solution of 1,3-bis(methylthio)-5-fluoro-1-pentene (12) (1.5 g, 8.32 mmol) in THF (32 mL) was added powdered calcium carbonate (2.5 g, 24.96 mmol). The mixture was stirred, and a solution of mercuric chloride (6.8 g, 24.96 mmol) in water (17 mL) and THF (56 mL) was added. A pale green to dark yellow color change was observed. The mixture was heated at 65-70 °C for 20 h. The reaction mixture was filtered with suction through a pad of celite. The filter cake was washed with a 1/1 pentane/ methylene chloride mixture. The filtrate was washed with saturated aqueous sodium chloride $(3 \times 50 \text{ mL})$. The combined aqueous solution was extracted with the 1/1 pentane/methylene chloride solution (2 \times 100 mL). The combined organic solution was dried (MgSO₄), filtered, and concentrated in vacuo at 25 °C. After almost all the solvent was removed, the residue was purified by silica gel column chromatography (90/10 hexanes/ethyl acetate) to provide 0.50 g (59% yield) of trans-5-fluoro-2-pentenal as a bright yellow liquid: $R_f 0.57 (20/80$ ethyl ether/chloroform); ¹H NMR (CDCl₃) δ 9.53 (d, J = 8.0Hz, 1H), 6.84 (m, 1H), 6.20 (m, 1H), 4.59 (dt, J = 6.0, 12.0 Hz), 2H), 2.77-2.67 (m, 2H); ¹³C NMR (CDCl₃) δ 193.42, 152.3 (d, J = 4.6 Hz), 134.7, 81.2 (d, J = 168.6 Hz), 33.3 (d, J = 20.6Hz); IR (CHCl₃) 3013, 2966, 1731, 1690, 1637, 1461, 1373, 1243, 1138 cm⁻¹; HRMS m/e calcd for C₅H₇FO 102.0481, found 102.0483.

To a 5 mL hydrolysis tube charged with tris(triphenylphosphine)rhodium chloride (0.034 g, 0.037 mmol) was added chlorodiisopropylsilane (0.861 mL, 5.044 mmol). The resulting mixture was stirred for 5 min. *trans*-5-Fluoro-2-pentenal (0.500 g, 4.89 mmol) was then added *via* cannula. The tube was sealed and stirred at 25 °C for 20 h and then heated at 90 °C for 4 h, after which time the reaction was complete by ¹H NMR analysis. Because of its instability, the enol silyl chloride **13** was used for the next reaction without further purification: ¹H NMR (CDCl₃) δ 6.34 (dt, J = 12.0, 1.2 Hz, 1H), 2.53–2.17 (m, 2H), 2.01 (m, 2H), 1.71 (m, 2H), 1.09 (d, J = 4.8 Hz, 6H), 1.07 (d, J = 4.8 Hz, 6H).

Pyrone Enol Silaketal (S)-14. To pyrone alcohol (S)-7 (1.00 g, 4.69 mmol) were added CH₂Cl₂ (10 mL) and triethylamine (1.3 mL, 9.39 mmol). The resulting dark brown solution was cooled to -78 °C and stirred for 5 min. Freshly prepared enol silyl chloride 13 (1.78 g, 7.04 mmol) was added via cannula. The dry ice bath was removed and the reaction mixture stirred at 25 °C until reaction was complete by TLC analysis (ca. 4 h). The reaction mixture was diluted with CH_{2} -Cl₂ (50 mL) and washed with water (20 mL). The aqueous wash was extracted with CH_2Cl_2 (2 \times 50 mL). The combined organic solution was dried over anhydrous MgSO₄, filtered, and evaporated under reduced pressure. The residue was subjected immediately to silica gel column chromatography (85/15 hexanes/ethyl acetate) to provide 1.12 g of pyrone enol ether silaketal (S)-14 in 56% yield as a bright yellow oil: R_f 0.44 (60/40 hexane/ethyl acetate); ¹H NMR (CDCl₃) δ 8.13 (dd, J = 2.25, 6.78 Hz, 1H), 7.67 (dd, J = 2.28, 5.04 Hz, 1H), 6.35 (m, 2H), 5.28 (m, 1H), 5.01 (dt, J = 7.53, 15.0 Hz, 1H), 4.53-4.34 (m, 2H), 3.89 (m, 2H), 2.04–1.64 (m, 6H), 1.37 (d, J =6.27 Hz, 3H), 1.03 (d, J = 2.4 Hz, 12H), 1.03-1.00 (m, 2H); IR $(CHCl_3)\, 3013,\, 2943,\, 2860,\, 1760,\, 1702,\, 1549,\, 1284,\, 1260,\, 1226,\,$ 1202, 1096 cm⁻¹. Because of its instability, silaketal (S)-14 was carried on immediately.

Bicyclic Lactones (-)-15. To 0.730 g (0.70 mmol) of pyrone enol ether (S)-14, 7 mL of anhydrous ethyl ether, and 14 mL of anhydrous toluene was added 0.463 g (2.04 mmol) of ZnBr₂ (98% purity) predried under a vacuum pump for at least 8 h. The reaction mixture was cooled to -10 °C and stirred until reaction was complete by TLC analysis (*ca.* 10 days). The reaction mixture was dissolved in ethyl acetate, filtered through a plug of silica gel, concentrated, and subjected to

silica gel column chromatography (90/10 hexanes/ethyl acetate) to afford 0.337 g of the cycloadduct (-)-15a in 46% yield, R_f 0.56 (60/40 hexane/ethyl acetate), along with 0.088 g of cycloadduct (-)-15b in 12% vield. (-)-15a: ¹H NMR (CDCl₃) δ 6.81 (m, 1H), 6.62 (dd, J = 5.2, 7.6 Hz, 1H), 5.53 (m, 1H), 5.01 (dt, J = 1.2, 4.8 Hz, 1H), 4.54 (m, 1H), 4.44 (s, 1H), 3.92 -3.82 (m, 2H), 1.84-1.77 (m, 6H), 1.37 (d, J = 6.4 Hz, 3H),1.28-1.20 (m, 1H), 1.06-1.03 (m, 6H), 0.94-0.92 (m, 9H); ¹³C NMR (CDCl₃) δ 168.34, 167.0, 130.6, 129.6, 83.4 (d, J = 165.5Hz), 76.2, 75.1, 70.6, 62.6, 60.1, 49.0, 37.8, 28.8 (d, J = 19.8Hz), 26.3 (d, J = 4.5 Hz), 20.6, 17.7, 17.4, 17.3, 17.2, 12.7, 10.5; ¹⁹F NMR (CDCl₃) δ -219.20; IR (CHCl₃) 2913, 2872, 1766, 1731, 1619, 1461, 1361, 1284, 1114, 1061, 1038 cm⁻¹; HRMS m/e (M⁺ - *i*-Pr) calcd for C₁₈H₂₆FO₆Si 385.1483, found 385.1489; $[\alpha]^{31}_{D}$ -20.5 (c = 8.2 mg/mL, CHCl₃). (-)-15b: ¹H NMR (CHCl₃) δ 6.86 (dt, J = 1.6, 7.6 Hz, 1H), 6.53 (dd, J = $4.8,\ 7.6\ Hz,\ 1H),\ 5.60{-}5.55\ (m,\ 1H),\ 5.07{-}5.05\ (m,\ 1H),\ 4.99$ (dd, J = 0.8, 7.2 Hz, 1H), 4.52-4.36 (m, 2H), 4.01-3.96 (m, 2H)1H), 3.86-3.81 (m, 1H), 2.48-2.41 (m, 1H), 1.85-1.83 (m, 6H), 1.36 (d, J = 6.4 Hz, 3H), 1.06–1.01 (m, 6H), 0.96–0.93 (m, 9H); ¹⁹F NMR (CDCl₃) δ -219.62; [α]³¹_D -5.13 (c = 28 mg/ mL, CHCl₃).

Bis-Silyl Ether 16. To adduct (-)-15a (0.337 g, 0.786 mmol) was added 35 mL of 5% HF in acetonitrile at 25 °C. After 60 min, the reaction mixture was neutralized with aqueous sodium bicarbonate and extracted with $CHCl_3$ (3 \times 100 mL). The combined organic phase was dried $(MgSO_4)$ and concentrated in vacuo. The residue was dissolved in 10 mL of N,N-dimethylformamide (DMF). To this solution were added tert-butyldimethylsilyl trifluoromethanesulfonate (TB-DMSOTf) (0.867 mL, 3.774 mmol) and 2.6-lutidine (0.366 mL, 3.145 mmol). The progress of the reaction was monitored by TLC (ca. ~ 8 h). After the reaction was complete, the reaction mixture was dumped into 50 mL of water and extracted with ethyl ether $(3 \times 100 \text{ mL})$. The combined ethereal solution was dried (MgSO₄), concentrated under reduced pressure, and purified by silica gel column chromatography (93/7 hexanes/ ethyl acetate) to afford 0.383 g (89% yield) of bis-silyl ether 16 as a colorless oil: $R_f 0.63$ (60/40 hexane/ethyl acetate); ¹H NMR (CHCl₃) δ 6.72 (dt, J = 1.6, 8.0 Hz, 1H), 6.58 (dd, J =5.2, 8.0 Hz, 1H), 5.17 (m, 1H), 5.04 (dt, J = 1.6, 5.2 Hz, 1H), 4.61-4.38 (m, 2H), 4.19 (bs, 1H), 3.72 (t, 2H), 2.12-1.61 (m, 7H), 1.37 (d, J = 6.4 Hz, 3H), 0.89 (s, 9H), 0.79 (s, 9H), 0.08 (s, 3H), 0.06 (s, 3H), 0.05 (s, 6H); $^{13}\mathrm{C}$ NMR (CDCl₃) δ 168.8, 166.6, 130.5, 130.0, 83.4 (d, J = 166.3 Hz), 75.6, 73.4, 71.3, 62.9, 59.6, 49.0, 39.0, 28.0 (d, J = 18.8 Hz), 25.9(3), 25.5(3), 20.1, 18.2 (d, J = 3.8 Hz), 17.8, -4.4, -5.1, -5.3, -5.4; IR (CHCl₃) 2931, 2849, 1760, 1737, 1619, 1467, 1355, 1284, 1255, 1185, 1114, 1091, 908, 838 cm⁻¹; HRMS m/e (M⁺ – t-Bu) calcd for $C_{23}H_{40}FO_6Si_2$ 487.2347, found 487.2352.

Cyclohexenol Ester (-)-17. To a 50 mL oven-dried roundbottomed flask charged with 155 mg (0.284 mmol) of the bissilyl ether 16 was added at 0 °C 8.12 mL (20 equiv) of lithium allyl oxide in allyl alcohol (0.7 M). (The solution of lithium allyl oxide was prepared thusly: 4.92 mL of n-BuLi (1.42 M in hexanes) was added dropwise to a -78 °C solution of allyl alcohol. After addition was complete, the reaction mixture was allowed to warm to rt and stirred for 30 min.) The reaction mixture was stirred at 0 °C for 4 h and at 25 °C for 12 h. The reaction mixture was quenched with saturated NH₄Cl, and the product mixture was extracted with CH_2Cl_2 (3 × 50 mL), dried over MgSO₄, concentrated in vacuo, and chromatographed (90/ 10 hexanes/ethyl acetate) to provide 45 mg of cyclohexenol ester (-)-17 in 40% yield: $R_f 0.60$ (60/40 hexane/ethyl acetate); ¹H NMR (CHCl₃) δ 6.87 (dd, J = 2.4, 5.2 Hz, 1H), 5.06 (m, 1H), 4.63 (d, J = 2.0 Hz, 1H), 4.49 (t, J = 6 Hz, 1H), 4.43-4.37 (m, 2H), 3.63 (m, 2H), 2.51 (dt, J = 5.60, 11.2 Hz, 2H), 2.14-2.06 (m, 2H), 1.91-1.85 (m, 2H), 1.77-1.70 (m, 1H), 1.28(d, J = 6.4 Hz, 3H), 0.88 (s, 3H), 0.87 (s, 9H), 0.86 (s, 9H),0.15 (s, 3H), 0.07 (s, 3H), 0.02 (s, 6H); $^{13}\mathrm{C}$ NMR (CDCl₃) δ 165.9, 139.1, 131.3, 84.1 (d, J = 164.8 Hz), 68.9, 67.7, 64.8, 59.5, 47.0, 39.2, 31.1, 29.1 (d, J = 19.8 Hz), 25.9(3), 25.8(3), 20.3, 19.1 (d, J = 5.3 Hz), 18.2, 18.0, -4.43, -4.87, -5.37, -5.42; IR (CHCl₃) 3613, 2943, 2919, 2849, 1702, 1467, 1249,

1067, 832 cm⁻¹; HRMS m/e (M⁺ - t-Bu) calcd for C₂₂H₄₂FO₅-Si₂ 461.2555, found 461.2560; $[\alpha]^{20}_{D}$ -12.5 (c = 7.5 mg/mL, CHCl₃).

Allylic Alcohol (-)-18. To 103 mg (0.198 mmol) of cyclohexenol ester (-)-17 in 3.0 mL of DMF were added TBDMSOTf (0.100 mL, 0.436 mmol) and 2,6-lutidine (0.046 mL, 0.397 mmol) at 25 °C. After 24 h at rt, the product mixture was diluted with ether, washed with water, dried (MgSO₄), filtered, and concentrated under reduced pressure. Crude protected ester 17 was dissolved in 1 mL of CH₂Cl₂ and 2 mL of anhydrous toluene and cooled to -78 °C. To this solution was added i-Bu₂AlH (0.494 mL, 1 M in toluene). After 40 min, only a trace of starting material was present. The reaction mixture was guenched at -78 °C with 3 mL of sodium potassium tartrate (2 M in H₂O), and the resulting solution was warmed to rt. After 1 h, the reaction mixture was extracted with ethyl ether $(3 \times 50 \text{ mL})$, filtered, dried over anhydrous MgSO₄, evaporated under reduced pressure, and subjected to silica gel column chromatography (93/7 hexanes/ethyl acetate) to provide 75 mg of allylic alcohol (-)-18 in 87% yield: $R_f 0.55$ (70/ 30 hexane/ethyl acetate); ¹H NMR (CDCl₃) 5.67 (m, 1H), 4.53-4.45 (m, 1H), 4.27 - 4.35 (m, 1H), 4.14 (d, J = 1.6 Hz, 1H), 4.06(s, 2H), 2.18 (dt, J = 5.2, 11.2 Hz, 1H), 2.00 (m, 1H), 1.82-1.68 (m, 5H), 0.90 (s, 9H), 0.88 (s, 9H), 0.124 (s, 3H), 0.119 (s, 3H), 0.05 (s, 6H); ¹³C NMR (CDCL₃) & 136.9, 125.0, 84.3 (d, J = 164.8 Hz), 70.2, 66.03, 65.1, 47.3, 31.1, 29.3 (d, J = 19.8Hz). 25.8(6), 19.7, 19.6, 18.0 (d, J = 7.5 Hz), -4.25, -4.61, -4.77, -4.84; IR (CHCl₃) 3613, 2943, 2919, 2849, 1466, 1249, 1213, 1096, 1044, 1005, 873 cm⁻¹; HRMS $m/e (M^+ - t-Bu)$ calcd for ${\rm C}_{18}{\rm H}_{36}{\rm FO}_{3}{\rm Si}_{2}$ 375.2187, found 375.2184; $[\alpha]^{20}{}_{D}$ –41 $(c = 14.4 \text{ mg/mL}, \text{CHCl}_3).$

Z-Dienoate (-)-19. A 25 mL oven-dried hydrolysis tube was charged with allylic alcohol (-)-18 (0.070 g, 0.162 mmol) and 1-(phenylsulfinyl)-2,2,2-triethoxyethane (0.225 g, 0.785 mmol),¹⁴ 2,4,6-trimethylbenzoic acid (TMBA) (0.003 g, 0.018 mmol) and 5 mL of anhydrous CH₂Cl₂. The tube was sealed and heated at 145 °C for 14 h. The product mixture was cooled to 25 °C and then filtered through a plug of silica gel, and the filter cake was washed with ether. The filtrate was evaporated under reduced pressure and the residue chromatographed (95/5 hexanes/ethyl acetate) to give 0.074 g of product as a mixture of E- and Z-dienoates. A 10 mL borosilicate test tube was charged with the E/Z dienoate mixture, 9-fluorenone (5.3 mg, 0.03 mmol), and 5 mL of tert-butyl methyl ether. The test tube was then placed in a 2 M aqueous solution of sodium orthovanadate (Na_3VO_4) and irradiated with a medium pressure mercury arc lamp for 20 h. The product mixture was filtered through a plug of Celite, and the filter cake was washed with CH₂Cl₂. The filtrate was evaporated in vacuo, and the residue was subjected to preparative TLC purification (75/25 hexanes/ethyl acetate) to afford 0.050 g of the Z-dienoate (-)-19 in 70% overall yield: $R_f 0.82$ (70/30 hexane/ethyl acetate); ¹H NMR (CDCl₃) & 5.62 (s, 1H), 5.18 (s, 1H), 5.07 (s, 1H), 4.48 (t, J = 6.0 Hz, 1H), 4.39 - 4.34 (m, 2H), 4.24 (m, 2H),4.16-4.06 (m, 2H), 2.36-2.27 (m, 2H), 1.75-1.52 (m, 6H), 1.23 (t, J = 7.2 Hz, 3H), 0.91 (s, 9H), 0.87 (s, 9H), 0.08 (s, 3H), 0.07(s, 6H), 0.05 (s, 3H); ¹³C NMR (CDCl₃) δ 165.8, 152.9, 128.3, 117.5, 112.3, 84.2 (d, J = 164.8 Hz), 74.3, 68.4, 59.7, 50.7, 44.228.6 (d, J = 19.8 Hz), 25.79(3), 25.75(3), 21.8, 19.7, 18.1 (d, J= 6.8 Hz), 14.2, -4.22, -4.61, -5.03, -5.08; IR (CHCl₃) 2943, 2919, 2849, 1713, 1637, 1467, 1249, 1173, 1091 cm⁻¹; HRMS m/e calcd for C₂₆H₄₉FO₄Si₂ 500.3153, found 500.3156; [α]²⁰_D $-40.2 (c = 12 \text{ mg/mL}, \text{CHCl}_3).$

Phosphine Oxide (+)-**20.** To Z-dienoate (-)-**19** (0.086 g, 0.172 mmol), CH₂Cl₂ (2 mL), and toluene (4 mL) was added *i*-Bu₂AlH (0.515 mL, 1.0 M in toluene) at -78 °C. After 1 h, the reaction was complete by TLC analysis (70/30 hexanes/ ethyl acetate, R_f 0.56). The reaction was quenched at -78 °C with 6 mL of 2 M sodium potassium tartrate solution. Ethyl acetate (10 mL) was added, and the mixture was allowed to warm to 25 °C and stirred for 1 h. The product mixture was extracted with ethyl acetate (3 × 50 mL). The combined organic solution was dried (MgSO₄), filtered, and concentrated *in vacuo* to provide crude allylic alcohol.

To a separate flask charged with 0.183 g (1.37 mmol) of N-chlorosuccinimide (NCS) and CH_2Cl_2 (4 mL) was added

0.106 mL (1.44 mmol) of dimethyl sulfide (DMS) at 0 °C (the reaction mixture immediately turned to a white turbid solution). The NCS-DMS complex was allowed to stir at 0 °C for 30 min and then cooled to -20 °C with a dry ice ethylene glycol bath. To the NCS-DMS complex was added the allylic alcohol in 2 mL of CH_2Cl_2 via cannula at -20 °C. The reaction mixture was stirred at -20 °C for 15 min and then warmed to 0 °C and stirred for another 30 min, after which time reaction was complete by TLC analysis (70/30 hexanes/ethyl acetate, R_f 0.75). The product mixture was dissolved with water (10 mL) and CH₂Cl₂ (50 mL). The layers were separated, and the aqueous phase was extracted with CH_2Cl_2 (3 imes50 mL). The combined organic solution was dried $(MgSO_4)$, filtered, and concentrated under reduced pressure. The residue was filtered through a plug of florisil with 80/20 hexanes/ethyl acetate and concentrated in vacuo to provide the allylic chloride as a tan oil.

A solution of this allylic chloride in 2 mL of THF was cooled to -78 °C, and potassium diphenylphosphide (0.5 M in THF) was added (~ 0.5 mL) until the red color persisted. The reaction mixture was stirred at -78 °C until the reaction was complete (ca. 2 h). The reaction was guenched with water (1 mL) at -78 °C, and the mixture was warmed to 25 °C and concentrated to a reduced volume ($\sim 2 \text{ mL}$). To this solution was added 10-15 drops of 30% H_2O_2 . After 20 min, to the reaction mixture were added water (5 mL) and CH_2Cl_2 (20 mL). The layers were separated, and the aqueous layer was extracted with CH_2Cl_2 (3 \times 50 mL). The combined organic solution was dried $(MgSO_4)$, filtered, and concentrated in vacuo. The residue was subjected to silica gel column chromatography (70/30 hexanes/ethyl acetate) to afford 60 mg of phosphine oxide (+)-20 as a white, viscous oil in 54% overall yield from Z- dienoate (-)-19: Rf 0.65 (75:25-hexane:ethyl acetate); ¹H NMR (CDCl₃) & 7.74-7.68 (m, 4H), 7.75-7.44 (m, 6H), 5.29 (m, 1H), 5.09 (s, 1H), 4.77 (d, J = 1.6 Hz, 1H), 4.40 (dt, J = 6.4, 47.6 Hz, 2H), 4.15-4.08 (m, 2H), 3.42-3.12 (m,)2H), 2.30-2.18 (m, 2H), 1.78-1.53 (m, 5H), 0.89 (s, 9H), 0.82 (s, 9H), 0.05 (s, 3H), 0.01 (s, 3H), 0.00 (s, 3H), -0.03 (s, 3H);¹³C NMR (CDCl₃) δ 146.2 (t, J = 2.3 Hz), 141.0 (d, J = 12.2Hz), 133.2 (d, J = 7.6 Hz), 132.3 (d, J = 8.3 Hz), 131.7, 130.9(t, J = 8.3 Hz), 128.5 (dd, J = 4.6, 11.5 Hz), 114.6 (d, J = 7.7 Hz)Hz), 111.9 (m), 84.2 (d, J = 164.8 Hz), 74.5, 68.4, 50.6, 43.3, 31.5, 30.8, 28.6 (d, J = 19.9 Hz), 25.8(6), 21.5 (d, J = 4.6 Hz),18.1 (d, J = 6.8 Hz), -4.29, -4.45, -4.93, -5.08; IR (CHCl₃)2943, 2931, 2895, 2849, 1467, 1431, 1390, 1355, 1249, 1167, 1114, 1091, 1067 cm⁻¹; HRMS m/e calcd for C₃₆H₅₆FO₃PSi₂ 642.3490, found 642.3483; $[\alpha]^{20}$ _D +5.5 (c = 5.7 mg/mL, CH₂-Cl₂).

Fluoro Vitamin D Analog (-)-5. Phosphine oxide (+)-20 (0.070 g, 0.108 mmol, 1.70 equiv, after being azeotropically dried three times with benzene and held under high vacuum for at least 30 h) was dissolved in 1 mL of freshly distilled anhydrous THF and the mixture cooled to -78 °C under argon. To this was added PhLi (0.080 mL, 0.115 mmol, 1.7 equiv, 1.48 M in ether) dropwise over a period of 5 min, during which time a deep red color change occurred and persisted. The mixture was allowed to stir for an additional 6-8 min. A precooled (-78 °C) solution of C,D-ring ketone (+)-21 (0.024 g, 0.068 mmol, 1.0 equiv, dried in exactly the same manner as for the phosphine oxide) dissolved in 1 mL of THF was added dropwise via cannula. The deep red mixture was stirred at -78 °C in the dark for 1.5 h, after which time the red color began to fade and analytical TLC (90/10 hexanes/ethyl acetate) indicated that reaction was complete. The reaction was then quenched at -78 °C with 0.5 mL of 2 N sodium potassium tartrate, followed by addition of dilute aqueous K_2CO_3 (2 mL). After being warmed to rt, the reaction mixture was extracted with ethyl acetate $(3 \times 20 \text{ mL})$. The combined organic solution was dried (MgSO₄), filtered, evaporated under reduced pressure, and chromatographed (eluent 95/5 hexanes/ethyl acetate) to provide 0.012 g (22% yield) of coupled product. To the coupled product in 0.5 mL of anhydrous THF were added solid n-Bu₄-NF (0.023 g, 0.089 mmol, 5.5 equiv) and 4 Å molecular sieves. The mixture was then heated at 60 °C for 1 h, after which time only a trace of starting material was present by TLC analysis (20/80 hexanes/ethyl acetate). The reaction mixture was filtered through a plug of Celite and immediately chromatographed rapidly (eluent 50/50 hexanes/ethyl acetate) to provide 0.0013 g (17% yield) of analog (-)-5 as an oil: R_f 0.62 (20/80 hexane/ethyl acetate); ¹H NMR (CDCl₃) δ 6.35 (d, J = 11.4 Hz, 1H), 6.02 (d, J = 11.0 Hz, 1H), 5.38 (t, J = 2.0 Hz, 1H), 5.03 (t, J = 2.0 Hz, 1H), 4.56-4.41 (m, 2H), 4.16-4.11 (m, 3H), 2.84-2.80 (m, 1H), 2.54-2.41 (m, 2H), 1.22 (s, 6H), 0.938 (d, J = 6.4 Hz, 3H), 0.549 (s, 3H); based on the ¹H NMR spectrum, the purity of this sample was judged to be at least 90%. Because of the small amount of (-)-5 available, only a poor quality ¹³C NMR spectrum could be obtained: ¹⁹F NMR (CDCl₃) δ -218.11 (m, 1F); IR (CHCl₃) 2943, 2872, 1725, 1602, 1461, 1373, 1261 cm⁻¹; UV (MeOH) λ_{max} 264 nm (ϵ 16 000), HRMS m/e calcd for C₃₀H₄₇FO₃ 474.3509, found 474.3505; [α]²⁰_D -32.2 (c 3.0 mg/mL, CHCl₃).

Silyl Enol Ether 22c. To a solution of 4 g (35.0 mmol) of ϵ -caprolactone in 100 mL of toluene at -78 °C was added dropwise, over 30 min, 36 mL (36 mmol) of a 1 M solution of *i*-Bu₂AlH. After addition was complete, the reaction was stirred for 1 h at -78 °C. The reaction was then quenched with 10 mL of a saturated solution of potassium sodium tartrate, diluted with 100 mL of EtOAc, and stirred for 30 min while it was allowed to warm to rt. The layers were separated, and the organic layer was washed with brine, dried over MgSO₄, filtered, and concentrated. Column chromotography provided 3 g (25.9 mmol, 74% yield) of the desired hydroxy aldehyde. This material was dissolved in 50 mL of CH_2Cl_2 . To this solution were added 3.6 mL (25.9 mmol) of Et_3N and approximately 20 mg of DMAP, followed by 5.44 mL (25.9 mmol) of chlorodiphenylmethylsilane. The reaction was allowed to run for 12 h, by which time a large amount of white solid had formed, signaling the completion of the reaction. Addition of 25 mL H₂O, followed by separation of the organic layer from the aqueous, drying the organic layer over $MgSO_4$, filtration, and concentration provided a yellow oil. This oil was purified by column chromatography to give 6 g (19.2 mmol, 55% yield from ϵ -caprolactone) of O-protected aldehyde: ¹H NMR (400 MHz, CDCl₃) δ 9.73 (s, 1H), 7.58 (m, 4H), 7.38 (m, 6H), 3.70 (t, J = 3.2 Hz, 2H), 2.38 (m, 2 H), 1.65–1.35 (m, 6H), 0.62 (s, 3H); IR (CHCl₃) 2731, 1725, 1619, 1584 cm⁻¹.

The above aldehyde was then diluted with 35 mL of benzene and 3.2 mL (23 mmol) of triethylamine. The resulting mixture was cooled to 0 °C, and 5.3 mL (23 mmol) of tert-butyldimeth-ylsilyl triflate was added. The reaction mixture was allowed to warm to rt and stirred for 1 h. The reaction was quenched with brine and the mixture diluted with Et_2O . The organic layer was separated, and the aqueous layer was washed with Et₂O. The combined organic layers were dried over MgSO₄, filtered, and concentrated. The crude product was purified by column chromatography (hexane) on silica gel that was slurrypacked with 1% triethylamine/hexane. The silylated vinylic ether 22c (3.6 g, 8.5 mmol, 65% yield) was a clear oil: ¹H NMR (400 MHz, CDCl₃) δ 7.58 (m, 4H), 7.38 (m, 6H), 6.15 (d, J =7.5 Hz, 1H), 4.41 (dt, J = 7.5, 1.3 Hz, 1H), 3.69 (t, J = 3.4 Hz, 2H), 2.05 (m, 2H), 1.57 (m, 2H), 1.37 (m, 2H), 0.91 (s, 9H), 0.61 (s, 3H), 0.11 (s, 6H); $^{13}{\rm C}$ NMR (100 MHz, CDCl₃) δ 138.7, 136.4, 134.3, 129.7, 127.9, 127.8, 110.6, 63.6, 32.4, 25.8, 23.5, -2.8, -5.2; IR (CHCl₃) 1648, 1472, 1431, 1255, 1114, 1084 cm⁻¹; HRMS m/e (M - t-Bu) calcd for C₂₁H₂₉O₂Si₂ 369.1706, found 369.1708.

Cycloadduct (\pm)-23c. A 12 cm piece of 3/8 in. heat shrinkable teflon tubing (Ace Glass catalog no. 12685-40) was sealed on one end with a glass dowel plug by using a heat gun. To this were added 0.50 g (3.24 mmol) of methyl 2-pyrone-3carboxylate, 4.15 g (9.70 mmol) of silvlated vinylic ether (\pm) -**22c**, 10 mg of barium carbonate, and 2 mL of dry CH_2Cl_2 . The open end of the tubing was then sealed in a similar fashion with a second glass dowel plug. This "sealed tube" was then pressurized at 10-11 Kbar at rt for 5 days. The reaction mixture was concentrated on a rotary evaporator, and the residue was purified by column chromatography (5% EtOAc/ hexane) to give 1.16 g (2.00 mmol, 62%) of the cycloadduct (\pm)-**23c** as a clear oil: $R_f 0.65$ (75/25 hexane/ethyl acetate); ¹H NMR (400 MHz, $CDCl_3$) δ 7.59 (m, 4H), 7.40 (m, 6H), 6.77 (dd, J = 5.0, 1.7 Hz, 1H), 6.44 (dd, J = 7.8, 5.0 Hz, 1H), 5.06 (ddd, J = 7.8, 4.7, 1.7 Hz, 1H), 4.78 (dd, J = 7.6, 1.0 Hz, 1H), 3.89

(s, 3H), 3.70 (t, J = 3.2 Hz, 2H), 2.34 (ddd, J = 7.6, 4.7, 1.0 Hz, 1H), 1.65–1.35 (m, 6H), 0.78 (s, 9H), 0.68 (s, 3H), 0.02 (s, 3H), 0.01 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 168.9, 167.5, 134.2(4), 133.9(2), 130.5, 129.7(2), 128.6, 127.8(4), 76.1, 69.4, 63.3, 62.9, 52.7, 44.8, 32.5, 27.3, 25.7, 23.6, 18.1, -3.2, -3.9, -5.0; IR (CHCl₃) 1760, 1742, 1619, 1584 cm⁻¹; HRMS m/e (M – t-Bu) calcd for $C_{28}H_{35}O_6Si_2$ 523.1972, found 523.1976. A diastereomeric cis-4,5-disubstituted cycloadduct was also isolated (25 mg, 0.09 mmol, 3% yield): R_f 0.60 (75/25 hexane/ethyl acetate); ¹H NMR (400 MHz, CDCl₃) δ 7.60 (m, 4H), 7.40 (m, 6H), 6.55 (dd, J = 7.8, 5.2 Hz, 1H), 6.40 (dd, J = 7.8, 2.0 Hz, 1H), 4.99 (dd, J = 4.3, 0.6 Hz, 1H), 4.32 (d, J = 8.6 Hz, 1H), 3.80 (s, 3H), 3.70 (t, J = 3.2 Hz, 2H), 1.90 (ddt, J = 7.8, 2.0, 1.0 Hz, 1H), 1.65–1.35 (m, 6H), 0.85 (s, 9H), 0.62 (s, 3H), 0.02 (s, 6H).

Alcohol (±)-24. To 931 mg (1.6 mmol) of cycloadduct (±)-23c in 7 mL of DMF were added 114 mg (1.7 mmol) of NaN_3 and 95 mg (1.7 mmol) of anhydrous NH_4Cl . The reaction was run at rt for 1 day. After completion, 1 mL of water was added and the reaction mixture was diluted with 10 mL of ethyl acetate. The layers were separated, the aqueous layer was washed with 10 mL of ethyl acetate, and the organic layers were combined, dried (MgSO₄), filtered, and concentrated. The reaction mixture was purified via short-path column chromotography (10 g of silica gel, 25-75% EtOAc/hexane) to give 384 mg (1 mmol, 62%) of the alcohol (\pm)-24 as a white solid: ¹H NMR (400 MHz, CDCl₃) δ 6.77 (dd, J = 5.0, 1.7 Hz, 1H), $6.44 \,(dd, J = 7.8, \, 5.0 \,\,\mathrm{Hz}, \, 1\mathrm{H}), \, 5.06 \,(ddd, J = 7.8, \, 4.7, \, 1.7 \,\,\mathrm{Hz},$ 1H), 4.78 (dd, J = 7.6, 1.0 Hz, 1H), 3.89 (s, 3H), 3.70 (t, J =3.2 Hz, 2H), 2.34 (ddd, J = 7.6, 4.7, 1.0 Hz, 1H), 1.65–1.35 (m, 6H), 0.78 (s, 9H), 0.02 (s, 3H), 0.01 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 169.1, 167.5, 130.4, 128.7, 76.7, 69.4, 63.2, 62.1, 52.7, 44.8, 32.6, 27.4, 25.6, 23.5, 18.0, -3.9, -5.0; IR (CHCl₃) 3542, 1754, 1742, 1619, 1472, 1458, 1431 cm⁻¹; HRMS m/e(M - t-Bu) calcd for $C_{15}H_{23}O_6Si$ 384.1968, found 384.1975.

Fluorinated Cycloadduct (±)-25. To 78 µL (0.59 mmol) of diethylaminosulfur trifluoride in 1 mL of CH_2Cl_2 at -78 °C was added dropwise via cannula 205 mg (0.53 mmol) of alcohol (\pm) -7 in 5 mL of CH₂Cl₂. The reaction mixture was warmed to 0 °C, and the reaction was followed by TLC till complete after 2 h. The reaction was then quenched with water and the mixture allowed to warm to rt. After it warmed, it was diluted with CH_2Cl_2 , washed with brine, dried (MgSO₄), and concentrated. The residue was purified by column chromotography (5 g of silica, 0-25% EtOAc/hexane) to give 119 mg (0.31 mmol, 58%) of the fluoride (±)-25 as a white solid: R_f 0.39 (75/25 hexane/ethyl acetate); ¹H NMR (400 MHz, CDCl₃) δ 6.79 (dd, J = 5.0, 1.7 Hz, 1H), 6.52 (dd, J = 7.8, 5.0 Hz, 1H), 5.13 (ddd, J = 7.8, 3.4, 1.7 Hz, 1H), 4.76 (dd, J = 7.2, 1.0 Hz, 1H), 4.45 (dt, J = 5.9 Hz, 41.5H), 3.88 (s, 3H), 2.41 (ddd, J =7.2, 3.4, 1.0 Hz, 1H), 1.75-1.42 (m, 6H), 0.80 (s, 9H), 0.05 (s, 3H), 0.02 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 168.9, 167.5, 130.7, 128.6, 83.5 ($J_{CF} = 164 \text{ Hz}$), 76.1, 69.4, 63.3, 52.8, 44.9, $30.5 (J_{CF} = 19.7 \text{ Hz}), 27.3, 25.7, 23.2, 18.1, -3.9, -4.9; {}^{19}\text{F NMR}$ $(375 \text{ MHz}, \text{ CCl}_3\text{F}) \delta$ -218.71; IR (CHCl₃) 1760, 1743, 1473, 1461, 1437, 1284, 1260.7, 1132, 1102, 1073, 1008 cm⁻¹; HRMS m/e (M – t-Bu) calcd for C₁₅H₂₂O₅FSi 329.1221, found 329.1224.

Mixed Malonate (\pm) -26. To 500 mg (0.80 mmol) of bicyclic lactone (\pm) -25 and 3 mL of CH₂Cl₂ at 0 °C was added dropwise, via syringe, 1.34 mL (1.34 mmol) of a freshly made 1.0 M solution of lithium allyl oxide in allyl alcohol. Reaction was complete by TLC after 2 h. The reaction was quenched with 2 mL of aqueous NH₄Cl and the mixture extracted with CH₂- Cl_2 . The organic layer was dried with $MgSO_4$, filtered, and concentrated on a rotary evaporator. The residue was purified by column chromatography (0-20% EtOAc/hexane) to give 383 mg (0.86 mmol, 77%) of the desired malonate (\pm)-26 as a clear oil: ¹H NMR (400 MHz, CDCl₃) δ 5.97 (s, 2H), 5.85 (ddt, J =16.2, 10.5, 4.9 Hz, 1H), 5.25 (ddd, J = 16.2, 5.3, 1.5 Hz, 2H), 7.6, 4.5, 1.5 Hz, 1H), 4.46 (dt, J = 41.5, 5.9 Hz, 2H), 4.07 (d, J= 8.6 Hz, 1H), 3.75 (s, 3H), 1.79-1.44 (m, 6H), 1.38 (s, 1H), $0.82~(s,~9H),~0.12~(s,~3H),~-0.03~(s,~3H);~^{13}C$ NMR (100 MHz, $CDCl_3$) δ 169.0, 168.0, 134.0, 131.1, 122.4, 118.4, 83.5 (J_{CF} = 164 Hz), 71.7, 68.5, 65.8, 61.3, 52.6, 45.9, $30.1 (J_{CF} = 19 \text{ Hz})$, 26.6, 25.9, 22.6, 18.2, -3.9, -4.5; IR (CHCl₃) 1735, 1473, **Conjugated Ester** (±)-27. A mixture of 380 mg (0.86 mmol) of allyl malonate (±)-26, 44 μ L (1.07 mmol) of formic acid, 154 μ L (1.11 mmol) of triethylamine, 18 mg (0.07 mmol) of triphenylphosphine, and 4 mg (0.02 mmol) of palladium(II) acetate in 2 mL of dioxane was sealed in a 5 mL hydrolysis tube and the tube heated at 100 °C for 12 h. After evaporation of dioxane, 1 N HCl (1mL) was added, and the mixture was extracted with CH₂Cl₂ (2 mL × 2). The organic solution was washed with saturated NaHCO₃, dried over MgSO₄, filtered, and concentrated. The oily residue was purified by column chromatography (0–10% EtOAc/hexane) to give 266 mg (0.74 mmol, 86%) of a free alcohol conjugated ester that was then O-protected.

To 266 mg (0.74 mmol) of this alcohol ester, 104 μ L (0.89 mmol) of 2.6-lutidine, and 2 mL of CH₂Cl₂ at 0 °C was added dropwise via syringe 204 μ L (0.89 mmol) of tert-butyldimethylsilyl triflate. The reaction was complete by TLC after 10 min. The reaction was quenched at 0 °C with 1 mL of water and the mixture allowed to warm to rt. Extraction with CH₂-Cl₂ followed by drying with MgSO₄, filtration, and concentration afforded a viscous oil which was purified by column chromatography (5% EtOAc/hexane), giving 310 mg (0.65 mmol, 88%) of ester (\pm) -27 as a clear oil: ¹H NMR (400 MHz, $CDCl_3$) δ 6.89 (t, J = 3.2 Hz, 1H), 4.75 (d, J = 2.6 Hz, 1H), 4.46 (dt, J = 47.3, 6.1 Hz, 2H), 4.01 (ddd, J = 9.1, 6.1, 2.9 Hz)1H), 3.73 (s, 3H), 2.63 (ddd, J = 19.7, 6.1, 4.7 Hz, 1H), 2.05(ddd, J = 19.7, 9.1, 3.2 Hz, 1H), 1.79-1.32 (m, 7H), 0.90 (s, 7H)9H), 0.82 (s, 9H), 0.12 (s, 3H), 0.06 (s, 6H), -0.09 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 166.9, 140.5, 133.0, 83.8 ($J_{CF} = 163$ Hz), 66.8, 65.1, 51.4, 47.4, 37.1, 30.6 ($J_{\rm CF} = 19$ Hz), 26.0(3), 25.8(3), 25.0, 22.5, 18.4, 18.0, -4.2, -4.4, -4.8, -5.6; IR (CHCl₃) 1713, 1655, 1472, 1461, 1437, 1390, 1255 cm⁻¹; HRMS m/e (M - t-Bu) calcd for C₂₀H₃₈O₄FSi₂ 417.2293, found 417.2289.

Allylic Alcohol (±)-28. To 300 mg (0.63 mmol) of cyclohexene ester (\pm) -27 in 2 mL of toluene at -78 °C was added dropwise via syringe 1.39 mL (1.39 mmol) of 1 M i-Bu₂AlH in toluene. The reaction was complete by TLC after 1 h. The reaction was quenched by addition at -78 °C of 2 mL of saturated potassium sodium tartrate followed by dilution with 5 mL of EtOAc. After the mixture was allowed to warm to rt and stirred for 0.5 h, two layers were visible. These were separated, and the aqueous layer was washed with EtOAc. The combined organic fractions were washed with water, and brine, dried over MgSO₄, filtered, and concentrated. The residue was purified by column chromatography (0-25% EtOAc/hexane) to give 262 mg (0.59 mmol, 94%) of allylic alcohol (±)-28 as a clear oil: $R_f 0.47$ (75/25 hexane/ethyl acetate); ¹H NMR (400 MHz, CDCl₃) δ 5.51 (t, J = 1.50 Hz, 1H), 4.80 (d, J = 2.6 Hz, 1H), 4.45 (dt, J = 47.3, 6.1 Hz, 2H), 4.22 (d, J = 11.9 Hz, 1H), 4.04 (dd, J = 2.4, 0.5 Hz, 1H), 3.90(d, J = 11.9 Hz, 1H), 2.36 (ddd, J = 19.7, 6.1, 4.7 Hz, 1H),1.93 (ddd, J = 19.7, 9.1, 3.2 Hz, 1H), 1.79-1.62 (m, 6H), 0.93(s, 9H), 0.89 (s, 9H), 0.14 (s, 3H), 0.11 (s, 3H), 0.06 (s, 3H), 0.05 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 137.8, 122.2, 83.8 $(J_{\rm CF} = 163 \text{ Hz}), 69.5, 68.1, 65.1, 46.3, 31.5, 30.6 (J_{\rm CF} = 19 \text{ Hz}),$ 25.8(3), 25.7(3), 24.5, 23.3, 17.9, -4.4, -4.9, -5.0(2); IR (CHCl₃) 3518, 1472, 1460, 1390, 1361, 1255, 1067 cm⁻¹; HRMS m/e (M - t-Bu) calcd for $C_{19}H_{38}O_3FSi_2$ 389.2344, found 389.2344.

Dienyl Ester (±)-29. To 262 mg (0.59 mmol) of allylic alcohol (±)-28 in a sealable hydrolysis tube were added 420 mg (1.47 mmol) of (triethylphenyl)sulfinyl orthoacetate, 1.0 mg of 2,4,6-trimethylbenzoic acid, and 2 mL of CH₂Cl₂. The tube was purged with argon and sealed and then heated at 110 °C for 12 h. The reaction mixture was concentrated. Crude ¹H NMR showed a >8:1 mixture of Z:E isomers. The crude product was purified via PTLC (10% EtOAc/hexane) to give 239 mg (0.46 mmol, 79%) of Z-dienoate (±)-29 as a clear oil: $R_f 0.8$ (75/25 hexane/ethyl acetate); ¹H NMR (400 MHz, CDCl₃) δ 5.62 (s, 1H), 5.19 (t, J = 1.5 Hz, 1H), 5.11 (t, J = 1.1 Hz, 1H), 4.52 (dd, J = 6.1, 1.1 Hz, 2H), 4.45 (dt, J = 47.3, 6.1 Hz, 2H), 4.11 (q, J = 7.2 Hz, 2H), 3.97 (dd, J = 5.8, 2.0 Hz, 1H), 2.48 (dd, J = 5.2, 1.4 Hz, 1H), 2.16 (dd, J = 6.1, 1.7 Hz, 1H), 1.70 (m, 1H), 1.58 (m, 2H), 1.50 (s, 1H), 1.25 (m, 2H), 1.23 (m,

2H), 0.91 (s, 9H), 0.88 (s, 9H), 0.83 (s, 9H), 0.06 (s, 6H), 0.05 (s, 3H), 0.04 (s, 3H); ^{13}C NMR (100 MHz, CDCl₃) δ 165.8, 153.1, 145.2, 117.3, 117.3, 83.9 ($J_{\rm CF}$ 163 Hz), 72.7, 70.2, 59.7, 50.5, 44.9, 32.8, 30.8 ($J_{\rm CF}$ 19 Hz), 25.8(3), 25.7(3), 25.1, 23.6, 18.2, 17.9, 14.1, -4.6, -4.8, -4.9, -5.2, -5.17; IR (CHCl₃) 1719, 1637, 1472, 1461, 1390, 1373, 1361, 1255, 1185, 1096, 1078 cm⁻¹; HRMS m/e calcd for $C_{27}H_{51}O_4FSi_2$ 514.3310, found 514.3313.

Phosphine Oxide (\pm) -30. The same procedure used for the conversion of dienoate (-)-19 into phosphine oxide (+)-20 was used here. Dienoate (\pm) -29 (170 mg, 0.33 mmol) gave a crude product that was purified by column chromatography (25-50% EtOAc/hexane) to give 80 mg (0.12 mmol, 37% from Z-dienoate (\pm) -29) of the phosphine oxide (\pm) -30 as a clear oil: Rf 0.43 (25/75 hexane/ethyl acetate); ¹H NMR (400 MHz, CDCl_3 δ 5.31 (dd, J = 14.9, 6.6 Hz, 1H), 5.12 (s, 1H), 4.77 (s, 1H), 4.42 (d, J = 3.5 Hz, 1H), 4.41 (dt, J = 47.3, 6.1 Hz, 2H), 3.87 (dd, J = 9.1, 5.4 Hz, 1H), 3.40 (ddd, J = 15.1, 8.9, 1.5 Hz)1H), 3.14 (ddd, J = 16.0, 9.8, 6.3 Hz, 1H), 2.4 (d, J = 3.7 Hz, 1H), 2.06 (dt, J = 14.2, 2.9 Hz, 1H), 1.68 (m, 1H), 1.53 (m, 2H), 1.43 (m, 2H), 1.35 (m, 2H), 0.88 (s, 9H), 0.82 (s, 9H), 0.03(s, 3H), 0.01 (s, 3H), -0.01 (s, 3H), -0.03 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 146.1 ($J_{CP} = 2$ Hz), 140.9 ($J_{CP} = 12$ Hz), 131.7, 131.6 ($J_{CP} = 3 \text{ Hz}$), 130.9 ($J_{CP} = 3 \text{ Hz}$), 128.5 ($J_{CP} = 11$ Hz), 114.4 ($J_{CP} = 8$ Hz), 111.3, 83.9 ($J_{CF} = 163$ Hz), 72.5, 70.2, 50.6, 42.4, 31.5, 30.8, 25.8(3), 25.6(3), 25.1, 23.4, 18.1 ($J_{CF} =$ 17 Hz), -4.6, -4.7, -4.9, -5.1; IR (CHCl₃) 1472, 1455, 1437, 1390, 1360, 1255, 1172, 1120, 1096, 1072 cm⁻¹; HRMS m/e calcd for C₃₇H₅₈O₃FPSi₂ 656.3646, found 656.3656.

2-Fluorobutyl analogs (-)-4 and (+)-4'. To 67 mg (0.10 mmol) of phosphine oxide (\pm) -30 in 1 mL of THF at -78 °C was added dropwise 68 μ l (0.10 mmol) of a 1.48 M solution of phenyllithium in THF. The resultant red solution was allowed to stir for 10 min. A solution of 27 mg (0.076 mmol) of C,Dring ketone (+)-21 in 1 mL of THF was added via cannula. The reaction was complete after 1.5 h by TLC following the disappearance of the C,D ring. The reaction was quenched by addition of 1 mL of 1/1 KHCO₃/2 M potassium sodium tartrate. The layers were separated, and the organic phase was washed with brine, dried over MgSO4, filtered, and concentrated. The crude product was rapidly purified by filtration through silica gel using 10% EtOAc/hexane as eluent to give 36 mg (0.045 mmol, 60% yield) of the coupled product. The silvl ether was cleaved by redissolving the product in 1 mL of THF with 10 mg of 4 Å molecular sieves, treating the mixture with 60 mg (0.23 mmol) of solid tetrabutylammonium fluoride, and heating the mixture at 60 °C. After 3.5 h, the reaction mixture was concentrated, diluted with EtOAc, passed through a plug of silica gel, and concentrated. Column chromotography gave 10 mg (0.02 mmol, 50% yield) of a 1/2.5 mixture of diastereomers (-)-4 and (+)-4', respectively. The diastereomers were separated and purified by reverse phase HPLC (20% H₂O/CH₃CN on a C-18 semipreparative column): Major product (+)-4': $[\alpha]^{25}_{D}$ +23.3 (c = 1.5 mg/mL, CH₂Cl₂); UV (CH₂Cl₂) λ_{max} 259 nm (ϵ 18 600); ¹H NMR (400 MHz, CDCl_3) δ 6.40 (d, J = 10.3 Hz, 1H), 5.99 (d, J = 10.3 Hz, 1H), 5.27 (s, 1H), 4.99 (d, J = 2.4 Hz, 1H), 4.48 (dt, J = 41.5, 5.9 Hz, 2H), 4.37 (d, J = 2.4 Hz, 1H), 3.88 (m, 1H), 2.82 (d, J =9.8 Hz, 1H), 2.67 (dd, J = 9.8, 3.2 Hz, 1H), 2.26 (dd, J = 10.2, 3.2 Hz, 1H), 1.98 (t, J = 7.5 Hz, 1H), 1.75–1.23 (m), 1.21 (s, 6H), 0.93 (d, J = 7.2 Hz, 3H), 0.53 (s, 3H); ¹⁹F NMR (375 MHz, CCl₃F) δ -218.65; ¹³C NMR (125 MHz, CDCl₃) δ 146.6, 143.4, 132.5, 124.8, 116.7, 113.5, 94.75, 83.9 ($J_{\rm CF}$ = 163 Hz), 73.4 ($J_{\rm CF}$ = 18 Hz), 71.1, 70.3, 56.3, 49.5, 45.9, 44.4, 42.9, 40.5, 36.1, 29.7, 29.2(2), 27.6, 26.2, 22.2, 20.7, 18.8 ($J_{CF} = 17 \text{ Hz}$); HRMS calcd for C₃₁H₅₁O₃F, 490.3822, found 490.3827. Minor product (-)-4: $[\alpha]^{25}_{D}$ -12.5 (c = 1.2 mg/mL, CH₂Cl₂); UV (CH₂Cl₂) λ_{max} 263 nm (ϵ 20 000); ¹H NMR (400 MHz, CDCl₃) δ 6.42 (d, J = 11.1 Hz, 1H), 6.03 (d, J = 11.1 Hz, 1H), 5.28 (s, 1H), 5.01 (s, 1H), 4.48 (dt, J = 41.5, 5.9 Hz, 2H), 4.35 (s, 1H), 3.85 (m, 1H), 2.82 (m, 1H), 2.65 (m, 1H), 2.26 (m, 1H), 1.98 (t, J = 7.5 Hz, 1H), 1.75-1.23 (m), 1.22 (s, 6H), 0.94 (d, J = 7.2 Hz, 3H), 0.55(s, 3H); ¹⁹F NMR (375 MHz, CCl₃F) δ -219.46; insufficient quantity available for ¹³C NMR; HRMS calcd for C₃₁H₅₁O₃F 490.3822, found 490.3822.

Antiproliferative and Cell Differentiation-Inducing Activity.¹ HL-60 cells were seeded at 3.0×10^5 cells/mL in 2.0 mL of RPMI1640 medium supplemented with 10% heatinactivated fetal calf serum and kanamycin (60 µg/mL, Sigma). 1,25-(OH)₂D₃ or its analogs dissolved in 2 µL of ethanol were added. After 4 days of culture in a humidified atmosphere of 5% CO₂ in air at 37 °C, cells were mixed with an equivolume of 0.4% tripan blue, and then the viable cell number was counted under a microscope. Cell differentiation-inducing activities were examined by morphological obervations (Gimsa stain).

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Supporting Information Available: Spectroscopic data of all compounds and X-ray analysis of (-)-10b (53 pages). This material is contained in libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.

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